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(54) **Method for synthesizing stable, single-stranded cDNA in eukaryotes by transfection with a vector containing a bacterial retron and the vector used therefor**

Verfahren zur Synthese von stabiler, einzelsträndiger cDNS in Eukaryoten durch Transfektion mit einem bakteriellen "Retron" enthaltenden Vektor und der verwendete Vektor

Méthode pour synthétiser un cDNA simple brin stable dans les eucaryotes par transfection avec un vecteur contenant un "retron" bactérien et le vecteur utilisable dans la méthode

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**Description****FIELD OF THE INVENTION**

The invention concerns the field of recombinant DNA. More particularly, the invention relates to an *in vivo* method of synthesis of stable single-stranded cDNA in eucaryotic cells by means of a bacterial retroelement called a retron. The invention also relates to new eucaryotic vectors carrying the necessary elements to produce the single-stranded DNA-RNA hybrid structures. Moreover, the invention relates to transfected eucaryotes, *e.g.*, yeast, plant cells and mammalian cells. Uses are described for the new products.

**BACKGROUND**

Gram-negative bacteria such as *Myxococcus xanthus*, *Stigmatella aurantiaca* and *Escherichia coli* have been found to contain a retroelement called a *retron*. In TIBS, 16, 18-21 (1991a), the authors report on a peculiar type of satellite DNA, named multicopy single-stranded DNA (msDNA). These molecules are characterized by a structure which comprises a single-stranded DNA branching out of an internal guanosine residue of a single stranded RNA molecule by a unique 2',5'-phosphodiester linkage. These molecules are thus single-stranded DNA-RNA hybrids. Reverse transcriptase is required for the synthesis of these msDNAs. In Ann. Rev. Microbiol., 45, 163-186 (1991b), the authors present a comprehensive review on msDNAs. Also see msDNA in Bacteria, Lampson *et al.*, *Progress in Nucleic Acid Research and Molecular Biology*, 60, 1-24.

The production of single-stranded cDNA by reverse transcriptase as a template is an obligatory step for RT-mediated transcription of retroelements. See *Retroelements*. See Weiner *et al.*, Ann. Rev. Biochem., 55, 631-661 (1986) for review. This includes integration of retroviruses into mammalian genomes, production of infectious retroviruses from pro-viruses integrated into genomes, retrotransposition of retroelements, and formation of pseudo genes in eucaryotic cells.

However, single-stranded cDNAs produced *in vivo* by RT have never been directly detected, probably because of their instability.

While the production of msDNAs in bacteria has been a most significant development, the *in vivo* production of single-stranded DNAs in eucaryotic cells, *e.g.*, yeast or higher eucaryotic cells like plant and mammalian cells, is of even greater interest. Eucaryotes have well-known advantages over procaryotes for producing target molecules. There is an important need to produce stable single-stranded DNA in a sufficient yield for numerous practical uses in research and in industry. This invention has made an important contribution in that respect in producing single-stranded RNA-DNA structures which are detectable, stable and useful.

**SUMMARY OF THE INVENTION**

in accordance with the invention, a fundamental finding has been made. It has been discovered that single-stranded DNAs which are stable can be produced *in vivo* in eucaryotic cells.

The invention relates to a method of producing a stable msDNA in a eukaryotic cell ; a DNA expression vector ; and a eukaryotic host cell transformed with said vector as defined in the claims and illustrated below.

Briefly described, the invention provides a method (or process) for producing *in vivo* stable, single-stranded DNAs in eucaryotic cells like yeasts or plant cells or mammalian cells. The method of the invention produces a single-stranded cDNA by means of a retroelement called a retron. The single-stranded DNA is produced as an integral part of a branched RNA-linked multicopy single-stranded DNA (msDNA) structure. These structures are stable, *i.e.*, detectable after production and isolation in spite of the fact that they are constituted of RNA and DNA, both single-stranded. By the method of the invention also provides such msDNAs which contain foreign DNA and RNA fragments in the DNA and RNA portions, respectively, of the RNA-DNA structure can be produced. Though different from the known bacterial msDNAs, these molecules are designated msDNAs or "modified" msDNAs, because they have the characteristics and unique features of msDNAs as described herein.

The method of the invention also employs retrons. Retrongs are genetic elements which contain the coding region *msr* for the msRNA and *msd* for the msdDNA of the msDNA molecule, respectively, and the gene for reverse transcriptase (RT). The retrongs have sequences which are different from known bacterial retrongs in that the non-coding region has been shortened, specifically the region between the transcriptional initiation site of the selected promoter and the initiation codon of the RT gene.

The invention also provides DNA expression vectors containing retrongs which differ from known bacterial retrongs in that, the RT gene is positioned upstream of the *msr*-*msd* region, in reverse relationship of that in bacterial retrongs. These new retrongs produce greater yields of msDNAs.

The method of the invention produces msDNAs which contain a foreign DNA fragment in their DNA portion, for

instance, a single-stranded fragment complementary to the mRNA of a particular target gene (antisense DNA) and thus, may be valuable tools to inhibit or change the expression of undesirable proteins. Similarly, the msDNA can also contain a foreign RNA fragment.

Further novel embodiments of the invention are eucaryotic hosts transformed with the DNA expression vector discussed above.

Various uses for the single-stranded RNA-DNA structures are described, but do not form part of the invention.

#### **DEPOSIT OF GENETIC MATERIAL**

Plasmid YEp521-M1 has been deposited with the American Type Culture Collection (ATCC) under Accession No. 74092.

Plasmid YEp521-M4 has been deposited with the ATCC under Accession No. 74093.

Plasmid YEp521-M5 has been deposited with the ATCC under Accession No. 74094.

#### **BRIEF DESCRIPTION OF THE FIGURES**

FIG. 1 illustrates the biosynthetic pathway of msDNA synthesis.

FIG. 2A shows the structure of the typical bacterial msDNAs. FIG. 2B shows the structure of msDNA-Ye117.

FIG. 3 shows the arrangement of genes in the retroelement responsible for the production of msDNAs.

FIG. 4 shows a comparison of the domain structures of various bacterial RTs.

FIG. 5 shows plasmids YEp51 and YEp52.

FIG. 6 shows the restriction map of the 11.6-kb *Eco*R1 fragment.

FIG. 7 shows a diagrammatic representation of plasmid PC1-1BPv4, YEp521-M1, YEp521-M2, YEp521-M3, YEp521-M4 and YEp521-M5.

FIG. 8A shows bands a and b of a sequence polyacrylamide gel for the production of msDNA-Ec67 and FIG. 8B shows a schematic representation of extension of the 3' end of msDNA by AMV-RT and RNase A treatment.

FIG. 9 shows Southern blot hybridization of msDNA-Ec67 produced in *S. cerevisiae*.

FIG. 10 shows a diagrammatic representation of plasmid YEp521-M5. Darkened region in the retron represents 50-bp antisense DNA for cdc28 (cloned into the *Xho*I site) inserted into the *msd* region of retron Ec67. Also shown is the 50-bp antisense DNA for cdc28.

#### **DETAILED DESCRIPTION OF THE FIGURES**

FIG. 1 Biosynthetic pathway of msDNA synthesis. The retron region consisting of the *msr*-*msd* region and the gene for reverse transcriptase (RT) is shown on the top of the Figure. Solid arrows indicate the locations of two sets of inverted repeats (a1 and a2, and b1 and b2). Open arrows indicate the genes for *msd*RNA (*msr*), msDNA (*msd*), and RT. The primary transcript is considered to encompass the upstream region of *msr* through the RT gene, which is shown by a thin line at step 1. The thick region in the RNA transcript corresponds to the final *msd*RNA. The branched G residue is circled, and the initiation codon for RT is also shown. On the folded RNA, a triangle indicates the 5' end processing site at the mismatching base. The dotted lines at steps 3 and 4 represent DNA strands.

FIG. 2 (A) Structures of hybrid DNA-RNA msDNA are shown as follows: Mx162, Mx65, Sa163, Ec107, Ec67, Ec86 and Ec73. (Ann. Rev. Microbiol., 45, 163-186 (1991)) (B) Structure of hybrid DNA-RNA msDNA-Ye117 is shown. The hatched zone represents the anti-sense DNA of cdc28.

FIG. 3 Arrangement of genes in the retron element responsible for the production of msDNA. A single-copy retroelement on the bacterial chromosome contains the region required for the production of msDNA. All known msDNA coding regions contain three genes organized in a similar manner, as shown in (A): A gene, *msd*, codes for the DNA strand of msDNA. A second gene (*msr*) is situated, 5' to 3', in the opposite direction and codes for the RNA strand of msDNA. A closely positioned ORF codes for the RT. Transcription of this region initiates at or near the 5' end of *msr* and extends beyond *msd* to include the ORF. A set of inverted repeat sequences, a1 and a2, is also conserved among msDNA coding regions (short arrows). The circled G corresponds to the residue in the RNA that will contain the 2', 5' branch linkage in msDNA (see also FIG. 8A). (B) For the *E. coli* retron Ec67, the region encoding msDNA is only a small part of a large element found on the chromosome (open bar). The junction of the Ec67 retron with the host chromosome is flanked by 26-base directly repeated chromosomal sequences, as shown by arrows. The Figure is not drawn to scale.

FIG. 4 Domain structures of various bacterial RTs. The regions with closed bars and with stippled bars represent the RT and RNase H domains, respectively.

FIG. 5 Yeast expression vectors YEp51 (7.3-kb) and YEp52 (6.6-kb). The structures of two yeast expression vectors are diagrammed. Both are composed of sequences from the yeast plasmid 2- $\mu$ m circle (smooth single line) spanning

REP3 ( \_\_\_\_\_ ) and the origin of replication ( \_\_\_\_\_ ), from the bacterial plasmid pBR322 (jagged single line) spanning the ColE1 origin of replication and the gene conferring ampicillin resistance, from the yeast genome spanning the gene LEU2 ( \_\_\_\_\_ ), and from the region 5' to the yeast GAL10 gene ( \_\_\_\_\_ ), extending from the Sau3A site at -495 from the transcription-initiation site to the Sall site present in plasmid pNN78-Δ4 at +13. A cloned gene inserted in YEp51 in the Sall, Sall-to-BamHI, Sall-to-HindIII, or Sall-to-BclI sites pointed labelled I in the Figure, terminating at a site in the 2-μm-circle sequences indicated by the blocked arrow (T). Similar transcription would be obtained with genes inserted in the HindIII or HindIII to BclI sites of YEp52. Restriction enzymes: R, EcoRI; H, HindIII; B, BamHI; S, Sall; P, PstI; Bc, BclI. See Broach et al., Experimental Manipulation of Gene Expression, Academic Press Inc., New York, 1983.

FIG. 6 Restriction map of the 11.6-kb EcoRI fragment. In the Cl-E map, the left-hand half (EcoRI to HindIII) was not mapped. In the ClEP5 map, the locations and the orientations of msDNA and msdRNA are indicated by a small arrow and an open arrow, respectively. A large solid arrow represents an ORF and its orientation. See Lampson et al., Science, 243, 1033-1038 (1989).

FIG. 7 Diagrammatic representation of plasmid PC1-1BPv4, YEp521-M1, -M2, -M3, -M4 and M-5. Diagrams show only the regions (shaded bars) inserted in the yeast vector, YEp521. These regions contain retron-Ec67 and restriction sites shown are only those which are used for the construction of plasmids. Short arrows with msr or msd are the locations and the orientations of msdRNA and msDNA. Long arrows with RT represent the gene for RT and its orientation. Thick arrows represent the GAL10 promoter and its orientation of transcription. Letters on top of bars are the sites of restriction enzymes: H, HindIII; Ba, BamHI; Pv, PvuII; B, BamHI; and S, SmaI.

FIG. 8 A sequence polyacrylamide gel of the production of msDNA-Ec67 in S. cerevisiae.

(A) Total RNA prepared from 0.9 ml of a late-log culture was used for detecting msDNA with AMV-RT as described herein below. The RT reaction mixture was subjected to electrophoresis on a 6% sequence-urea-gel. An aliquot of the reaction mixture was treated with RNase A prior to gel electrophoresis. Lanes 1 and 2 (G and C lanes, respectively) are DNA sequence ladders of pUC19 sequenced by chain termination method (Sanger et al., Proc. Natl. Acad. Sci. USA, 74, 5463-5467 (1977)) for size marks; lane 3, the AMV-RT products with total RNA from yeast cells harboring YEp521-M1; lane 4, the same sample as lane 3 except that it was treated with RNase A prior to gel electrophoresis; lane 5, the AMV-RT products with total RNA from yeast cell harboring YEp521. The sample was treated with RNase A. Lane 6 is an MspI digest of pBR322 labeled with [ $\gamma$ -<sup>32</sup>P]dCTP with the Klenow fragment of DNA polymerase 1. Numbers at the right-hand side indicate fragment sizes in base pairs and arrows with letters indicate positions of msDNA.

(B) Schematic representation of extension of the 3' end of msDNA-Ec67 by AMV-RT and RNase A treatment.

FIG. 9 Southern blot hybridization of msDNA-Ec67 produced in S. cerevisiae.

(A) Total RNA fractions prepared from a 2.5 ml culture of yeast cells harboring YEp521-M1 (lane 1), and YEp521-M2 (lane 2) and from E. coli CL83 harboring pCL-1EP5c (lane 3) were used. After blotted to the nylon membrane filter, msDNA-Ec67 was detected with the nick-translated 140-bp msr-msd DNA fragment as a probe. An arrowhead indicates the position of msDNA-Ec67.

(B) Production of msDNA-Ec67 in S. cerevisiae harboring YEp521-M1, -M3, and -M4. Total RNA fractions prepared from a 2.5 ml culture of yeast cells harboring YEp521-M1 (lane 3), -M3 (lane 2), and -M4 (lane 1) were used for Southern blot hybridization as described in (A). An arrowhead indicates the position of msDNA-Ec67.

FIG. 10 YEp521-M5 was constructed from YEp521-M4 by inserting into the msd region an XhoI site and into that site, cloning a 50-bp extraneous (foreign) dsDNA fragment which is complementary to mRNA of cdc28; step 1. The XhoI site was added into the msd region of YEp521-M4 by PCR. This construct was then digested by XhoI; then the antisense DNA was ligated into the msd region of retron Ec67; step 2. This plasmid was transformed into yeast (SP-1) and the subsequently expressed msDNA designated herein as msDNA-Ye117. This is a novel structure.

#### DETAILED DESCRIPTION OF ILLUSTRATIVE EMBODIMENTS OF THE INVENTION

In accordance with this invention, it has been discovered that a transfected yeast, Saccharomyces cerevisiae, produces a genetic structure, described as a synthesized, branched RNA-linked multicopy single-stranded DNA (msDNA). One such msDNA produced was msDNA-Ec67. msDNA-Ec67 was synthesized from retron-Ec67.

The production of other representative msDNAs is described.

Several msDNAs have been described in the literature. Some of these are the following: Mx162 (Dhundale et al., Cell, 51, 1105-1112, 1987); Mx65 (Dhundale et al., J. Biol. Chem., 263, 9055-9058, 1988b); Sa163 (Furuichi et al., Cell, 48, 47-52, 1987a and Furuichi et al., Cell, 48, 55-62, 1987b); Ec67 (Lampson et al., Science, 243, 1033-1038, 1989b); Ec86 (Lim and Maas, Cell, 56, 891-904, 1989); Ec73 (Sun et al., J. Bacteriol 173, 4171-4181, 1991); Ec107

(Herzer et al., *Mol. Microbiol.*, submitted, August 1991); msDNA from *E. coli* B (Lim and Maas, *Cell*, 56, 891-904, 1989).

The msDNAs are often referred to in the literature by a numeral preceded by a suffix indicating a host origin. For instance, "Mx" referring to *Myxococcus xanthus*, and "Ec", referring to *E. coli* and "Sa" to *Stigmatella aurantiaca*.

msDNAs are unique molecules which in spite of extensive diversity, share similar structural features. Generically, msDNA may be described as being a molecule which comprises a branched RNA which is covalently linked to a single-stranded DNA by a 2',5'-phosphodiester bond between the 2'-OH group of an internal rG residue and the 5'-phosphate of the DNA molecule, and which RNA is non-covalently linked to the DNA by base pairing between the complementary 3' ends of the RNA and DNA molecules, which RNA and DNA form the stable stem-loop secondary structures. The msDNA molecule is encoded by a primary RNA transcript, pre-msDNA, which contains an open-reading frame (ORF) downstream of the *msr* locus encoding a polypeptide which has sequence similarity with retroviral RTs and a highly conserved sequence common to RTs.

The pre-msDNA may alternatively contain its ORF upstream of the *msr* locus, in which event the retron will be of like construction.

In FIG. 2 which shows typical msDNAs, the RNA portion of the molecule is shown "boxed"; the balance of the structure being the ssDNA portion.

It will be noted that the molecules all show a branched rG residue, a DNA-RNA hybrid at the 3' ends of the msDNA and msdRNA, and a stem-loop structure in the RNA and DNA strands. The branching ribonucleotide, G, is circled and the 2',5'-phosphodiester linkage to the first deoxynucleotide is indicated.

Retrons: a retron is a small genetic element to date found to be of 1.3 to 2.5-kb in length constituted of an *msr-msd* region and by the gene for encoding reverse transcriptase (RT). The coding region for msDNA is indicated by "*msd*"; the coding region for msdRNA is indicated by "*msr*".

A comparison of all known msDNA coding regions reveal that this locus contains three genes organized in a similar manner (see FIG.3). A gene called *msd* codes for the DNA portion of msDNA. A second gene, *msr*, is situated 5' to 3', in the opposite orientation of *msd*, and codes for the RNA chain. Thus the genes *msd* and *msr* are convergently oriented so that their respective 3' ends overlap by several bases.

This overlap is equivalent to the H-bonded DNA-RNA structure formed by the overlapping 3' ends of the RNA and DNA strands in the msDNA molecule. For Mx162, the overlapping *msd-msr* genes, like the hybrid structure of the msDNA they produce, comprise 8 base pairs. See Table I for typical overlap lengths of various msDNAs.

Determination of the nucleotide sequence in the vicinity of the *msd-msr* genes revealed a closely-linked open reading frame (ORF). This ORF is located immediately upstream from *msd*, but is transcribed in the same direction as *msr* (as shown in FIG. 7). The initiation codon of the ORF is situated as close as 19 basepairs from the start of the *msd* gene for the Ec86 retron of *E. coli* B, but as much as 77 base-pairs for the Mx162 retron of *M. xanthus*.

Another conserved feature of the chromosomal locus that codes for msDNA is a set of inverted repeat sequences, designated a1 and a2. Sequence a1 is located just upstream from the start of the *msd* gene, while sequence a2 is positioned immediately 5' to the G residue in the *msr* gene that forms the 2', 5' branch linkage in the msDNA molecule (FIG. 4). The inverted repeats display a large degree of nucleotide sequence diversity among the different known loci encoding msDNA, as well as differences in size. For example, the inverted repeats (a1 and a2) found in the retron locus encoding Mx162 are 34 nucleotides long, while the inverted repeats for the Ec86 retron of *E. coli* B are only 12 bases in size. Despite their diversity, these repeat sequences are located in the same positions (as shown in FIG. 3) for all known loci encoding msDNA. As discussed in more detail below, the position of these inverted repeat sequences is critical to the synthesis of msDNA.

It will be helpful to refer to the above discussion when the aspects are discussed which provide for an inversion in the organization (position inversion) of the RT gene with respect to the *msr-msd* coding region, and in the discussion of shortening the non-coding region between the transcriptional initiation site and the initiation codon AUG of the RT gene.

The promoter for the *msr-msd* region is upstream of *msr*. Transcription is from left to right, encompassing the entire region including the RT gene. As described in further detail hereinafter, the replicating vehicle for transfecting the eucaryote host may harbor one promoter for the *msr-msd* and the RT, or it may contain two promoters, one for the *msr-msd* region and the other for the RT.

In accordance with the invention retons may be constructed to yield an msDNA which differs from the typical msDNAs by features other than the common, conserved and characteristic features of msDNAs described above. For instance, it is not excluded that the length and/or location of the set of IRs a1 and a2 in the retons be varied providing they remain in the same location discussed above. Thus, the size and/or location of the loops in the stems of the msDNA can be varied.

Further, it is not excluded that the extent of the the overlap of the base pairing of the 3' ends of DNA and RNA in the msDNAs be influenced (increased or decreased) by appropriate manipulations. Whether such variations will be desirable will depend on the ultimate utility proposed for these msDNAs.

General Features of msDNAs. Table I is a summary of the structure of representative retons.

Reverse Transcriptase (RT). The domain structures of bacterial RTs of representative retrons are shown in FIG. 4.

The RT gene is normally located downstream from the *msr-msd* region. In the new retrons which differ from the bacterial retrons, their relative positions are reversed, the *msr-msd* region is located downstream of the RT gene.

The biosynthesis of msDNAs has been described by Inouye & Inouye, *Ann. Rev. Microbiol.*, **45**, 163-186 (1991b) and Herzer *et al.*, *Mol. Microbiol.*, submitted, August 1991. A schematic of the synthesis is shown in FIG. 1. A primary transcript (pre-msdRNA) is considered to encompass the upstream region of *msr* through the RT gene, which is by reference to FIG. 1, shown by a thin line at step 1. The thick region in the RNA transcript corresponds to the final msdRNA. The branched G residue is circled, and the initiation codon for RT is also shown. On the folded RNA, a triangle indicates the 5' end processing site at the mismatching base. The dotted lines at steps 3 and 4 represent DNA strands.

In summary, the primary transcript from the *msr-msd* region is believed to serve not only as a template but also as a primer to produce the msDNA. Synthesis of msDNA is primed from an internal rG residue of the RNA transcript using its 2'-OH group. Thus, msDNA is branched out from this rG residue by a 2'-5'-phosphodiester linkage.

There will be described hereinafter the transformation of yeast cells harboring plasmids which contain a retron (which includes the RT gene) for expression of the desired msDNAs. The description is of a best mode to date to express msDNA-Ec67 from its retron, Ec67.

1. Synthesis of msDNA-Ec67. For the expression of msDNA-Ec67, plasmid YEp52 was used. Plasmid YEp521 was constructed by introducing the multiple cloning sites of pUC19 (Yanisch-Perron *et al.*, *Gene*, **33**, 103-119, 1985) into YEp52, which was designed to obtain high-level, inducible expression of a cloned gene under the *GAL10* promoter in yeast. YEp52 contains the ColE1 origin of replication (OR), a promoter of the *GAL10* gene, *LEU2*, the 2 $\mu$ -circle origin of replication, and the 2- $\mu$  circle REP3 locus (SEE FIG. 5; See Broach *et al.*, *Experimental Manipulation of Gene Expression*, Academic Press Inc., New York, 1983).

Retron-Ec67 was prepared from plasmid pCL-1BPv4 in which the 4-kb *BalI*-*PvuII* fragment (DNA from fragment from the *BalI* to 2nd *PvuII* site from the left end of the map depicted in FIG. 5 was cloned into the *HincII* site of pUC9. *E. coli* harboring this plasmid produces msDNA-Ec67 (Lampson *et al.*, *Science*, **243**, 1033-1038, 1989).

A total RNA fraction was prepared from cells transfected with pC1-1EP5c; pC1-1EP5c contains the 5-kb *PstI*(a)-*EcoRI* fragment encompassing the entire 4-kb *BalI*-*PvuII* sequence in PC1-1BPv4. See FIG. 7.

The construction of plasmid YEp521 proceeded as follows. The DNA fragment containing the pUC19 multiple cloning sites were isolated by digestion of pUC19 with *EcoRI*, the cleaved ends were filled in with the Klenow fragment of DNA polymerase 1, and then digested with *HindIII*. The resulting 54-bp fragment was cloned into YEp52 by replacing a fragment between the *BclI* (filled in with the Klenow fragment) and *HindIII* sites, resulting in YEp521.

YEp521, thus constructed, contains the multiple cloning sites from pUC19, except for *EcoRI*, downstream of the *GAL10* promoter.

The 4-kb *HindIII*-*BamHI* fragment from pC1-1BPv4 (see FIG. 7) was cloned into the *HindIII* and *BamHI* sites of YEp521.

As a result, the *msr-msd* region and the RT gene of retron-Ec67 were placed downstream of the *GAL10* promoter. This plasmid is designated YEp521-M1.

Plasmid YEp521-M1 is illustrated in FIG. 7. The shade bars are the regions inserted in yeast vector, YEp521.

It will be noted that the RT of retron-Ec67 gene is located behind (downstream) the *msr-msd* region.

2. Production of msDNA in transformed yeast. A yeast strain (SP1: a *ura3, leu2, trp1, his3, ade8, can'*, *gal2*) was used. Transformation of the yeast cells was carried out by the lithium acetate method of (Ito *et al.*, *J. Bacteriol.*, **153**, 163-168, 1983). Yeast culturing was carried out as described below. msDNA was produced and was detected by extending the 3' end of msDNA by avian myeloblastosis virus reverse transcriptase (AMV-RT). This yielded a main product of 117 nucleotides. Treatment of this product with ribonuclease A resulted in a DNA of 105 nucleotides. These results are in good agreement with the structure of msDNA-Ec67 (see Lampson *et al.*, *Science*, **243**, 1033-1038, 1989). The production of msDNA-Ec67 was further conformed by Southern blot hybridization.

To determine whether the production of msDNA-Ec67 could occur in yeast without the RT genes from retron Ec67, the following work was performed.

An RNA preparation from cells harboring YEp521-M2 only containing the *msr-msd* region under the *GAL10* promoter, and described herein above (see FIG. 7) was analyzed by Southern blot hybridization. As shown in lane 2, FIG. 9, no band corresponding to msDNA-Ec67 was detected, indicating that the RT gene from retron-Ec67 is essential for the msDNA synthesis in yeast cells.

In a similar manner, CHO cells can be transformed using known strategies and techniques. The same could be done with HeLa cells or other vertebrate mammalian cells.

Gene Rearrangement in Retrons. An important finding in connection with the invention is that the yield of msDNA in transfected yeast cells is significantly improved by means which cause, it is believed, an increase in production of RT. One such strategy is to reduce by as many as possible the numbers of the AUG codons between the transcriptional initiation site of the *GAL10* promoter (or any other promoter used for that purpose) and the initiation codon of the RT

gene. Best results were obtained when a portion of the 5' end non-coding region containing initiation translation codons AUG is deleted, but for the first AUG codon in closest proximity to the 5' end.

Thus, it was found that a significant portion of the 5' end of the non-coding region was not essential to production of msDNA in yeast cells. Deletion of a portion of the nucleotide sequence containing the AUG codons significantly improved the yield of msDNA production.

Specifically in YEp521-M1 (see FIG. 7), there are 417 -bp from the 5' end HindIII site to the initiation codon GAA for RT. (See FIG. 7 in Lampson *et al.*, *Science*, 243, 1033-1038, 1989). A deletion of the 240-bp sequence upstream of the msr gene from the left hand most HindIII site to immediately upstream of msr of YEp521-M1 (see FIG. 7) was carried out.

For this purpose, the fragment of 140-bp msr-msd (including 5 extra bases upstream of msd and 18 extra bases at the 3' end of the msr-msd region upstream of msd) (after PCR amplification) and the 1.8-kb RT gene (including 8 bases upstream) of the initiation codon of the RT gene (also after PCR amplification) (and 4 bases downstream of the termination codon), were cloned into the HindIII and BamHI sites of YEp521-M1 yielding YEp521-M3 (see FIG. 7). The yield of msDNA-Ec67 in transfected yeast with YEp521-M3 was shown to be significantly increased, as discussed below.

Another important finding made to substantially increase the yield of msDNAs in yeast is to transpose the position of the RT gene with respect to the msr-msd region. In bacterial retransons, the msr-msd region is in front of the RT gene; when the RT was moved upstream of the msr-msd region, a further increase in yield of msDNA was observed. This was accomplished as follows.

Since the msr-msd region of YEp521-M3 still contains 3 AUG codons, YEp521-M4 (see FIG. 7) was constructed, in which the order of the RT gene and the msr-msd region was reversed, i.e., the msr-msd region being positioned after the RT gene. In YEp521-M4, there is only one AUG codon between the left hand-most HindIII site and the BamHI site (see FIG. 7), which exists in the multiple cloning sites of PUC19 (Yabisch-Perron *et al.*, *Gene*, 33, 103-119, 1985). This AUG codon is terminated by a termination codon, UAG after 5 codons. The initiation codon, GAA, for the RT gene was placed 6 codons after the termination codon in the same reading frame.

YEp-M5 was then constructed from YEp521-M4 by adding 50-bp antisense DNA for *cdc28* into the region coding for msd (described in further detail below). Therefore, two plasmids were constructed in which the order of the RT gene and the msr-msd region was reversed.

The yield of msDNA-Ec67 in yeast cells transfected with YEp521-M4 was compared with that of yeast cells transfected with YEp521-M3. YEp521-M4 brought about a further increase (about 8 fold) of yield over YEp521-M3.

It has been reported that a ribosomal subunit (carrying Met-tRNA<sup>met</sup> and various initiation factors) binds initially at the 5' end of mRNA and then scans through the mRNA stopping and then initiates translation at the first AUG codon in a favorable context (Kozak, *J. Cell Biology*, 108, 229-241, 1989). From a recent survey of 699 vertebrate mRNAs, **GCCGCCACCAUG** emerges as the consensus sequence for initiation of translation in higher eucaryotes (Boeke *et al.*, *Cell*, 40, 491-500, 1985). The survey reports the study of the 5' non-coding sequences of the 699 vertebrate mRNAs (all sequences to which access could be had in the literature). The mRNA source of the vertebrates included human (muscle, skeletal, liver, intestinal, etc.), bovine, rat and others. Also in yeasts, AUG was reported to be the consensus sequence for initiation of translation. (Hamilton *et al.*, *Nucl. Acids Res.*, 15, 3581-3583, 1987) It is noted that Kozak, *J. Cell Biology* 108, 229241, 1989 also reported variations and exceptions to the more general rule described above. For instance, there are reported cases where initiation is not restricted to the first AUG codon, which therefore is not used exclusively, but includes other AUG codons in the vicinity of the 5' end. Further, inactivating the first AUG codon closest to the 5' end, allowed ribosomes to initiate translation at another codon (UUG).

As described herein above, the location of the initiation codon of the ORF for various msDNAs can vary (e.g., 19-bp from the start of the msd gene for Ec86 retron and 77-bp for the Mx162 retron). Thus, one skilled in the art can adjust the length of the excised non-coding region of the retron when the above strategy is followed.

The finding in connection with the invention described above, namely, that a significant improvement in yield of msDNAs takes place when AUG codons between the transcriptional site of the GAL10 promoter and the initiation codon of the RT gene are deleted, but for the one AUG codon closest to the 5' end which is preserved, is therefore consistent with the above-discussed literature reports. Accordingly, this finding made in accordance with the invention with respect to the production of msDNAs is not intended to be limited to yeast, but can reasonably be predicted to apply to other msDNA-producing transfected eucaryotes, in particular higher eucaryotes like mammalian cells, e.g., HeLa cells, CHO, COS-1 cells and others.

The same observation can be made regarding the position of the RT gene upstream of the msr-msd region. This finding too is believed to have general applicability to the production of msDNAs in eucaryotes, as noted above. It is believed that these described strategies may contribute to an increase in RT and ultimately in yield of msDNAs.

It will be apparent to one skilled in the art that the two strategies described (deletion of AUG codons and inversion of the respective positions of the RT gene and the msr-msd region, do not have to be performed together (as shown with respect to YEp521-M4), which is a best mode to date. For instance, the strategy may be performed without the

deletion strategy, and vice-versa.

The msDNAs which are synthesized from these new retrons are also new.

As has been noted herein, it is not necessary that one promoter for the RT gene and the *msr-msd* region be used. More than one can be used, one for the RT and one for the *msr-msd* region. When it is desired to use two promoters, either one or both of the strategies to increase RT production namely the inversion and/or deletion strategy can also be used, as will be apparent to one skilled in the art.

It is noteworthy that the DNA sequences, which contain these unique retrons (due to the deletions and/or position inversion) and which encode the new msDNAs, are new when compared to known bacterial retrons. So are the replicating vehicles carrying these retrons and the transfected eucaryotes harboring these vehicles. They provide effective means to produce new single-stranded DNA in eucaryotes in improved yields.

It is to be noted also that the two above-described strategies which have been discussed with respect to eucaryotes are applicable to msDNAs produced from modified retrons in procaryotes.

The invention has been illustrated with an illustrative retron, Ec67. However, by a similar procedure, yeast can be made to produce other msDNAs. For instance, in a similar manner, retron Ec73 can be used to transform yeast strain SP1 to produce msDNA-Ec73.

Likewise, a similar procedure can be followed to transform and produce msDNA-Mx65 in yeast from the necessary retron elements. See Dhundale *et al.*, JBC, 263, 9055-9058, 1988. Its ORF codes for 427 amino acid residues.

If it is desired to produce msDNA-Mx162 in yeast, the appropriate DNA fragment containing retron Mx162 can be prepared from a 17.5-kb *SalI* fragment which is disclosed in Yee *et al.*, Cell, 38, 203-209, 1984. Its ORF provide for 485 amino acid residues.

For the expression of msDNA-Ec107, a similar strategy may be followed. The retron is a 1.3-kb DNA fragment of which the 34-bp intergenic sequence between *pyrE* and *tkk* (in FIG. 4) is deleted. The retron contains an ORF coding for 319 amino acid residues (from base 396 to 1352 in FIG. 2). The reference to Figures made hereinabove is to Dhundale *et al.*, Cell, 51, 1105, 1987. This retron is the smallest yet found in bacteria.

The retron for Sa163 was determined to be contained in a 480-bp DNA fragment encompassing the *msd* and *msr* regions (Furuichi *et al.*).

The retron for Ec73 was determined to be contained in a 3.5-kb *salI*(b)-*EcoRI*(c) fragment (see FIG. 1A of Sun *et al.*). For details on Ec73, see below.

The retron for Ec86 was determined to be contained in a 3.5-kb *PstI* fragment (Lim and Maas, Cell, 56, 891-904, 1989).

Likewise, from retrons Sa163, Ec86 and Ec73, the corresponding msDNAs, msDNA-Sa163, msDNA-Ec86 and msDNA-Ec73 may be produced in transfected yeast cells. If plant or mammalian vertebrate cells are used, appropriate manipulations and strategies will be followed.

Similar techniques may be followed to express other msDNAs known or yet to be found or to be synthesized from their respective retrons. All of these retrons are expected to contain the elements necessary to synthesize the unique features of msDNAs, as is described herein.

Thus, in general retrons containing the essential features described herein are useful to produce *in vivo* in eucaryotes the stable (not degraded) msDNAs having the conserved and characteristic features described herein.

msDNA-Ec73 is synthesized from retron Ec73 which is described by Sun *et al.*, Journal of Bacteriology, 173, 4171-4181, 1991.

FIG. 2 therein shows the nucleotide sequence capable of synthesizing msDNA-Ec73, a 3.5-kb S(b)-E(c) fragment. It was determined that the first ATG codon at position 11,544 is the initiation for the necessary RT gene and the ORF for the RT is of 316 residues.

It is to be noted that in all retrons known to date, the RT gene is located at 20 to 77-bp upstream of the *msd* gene (downstream of the *msd* gene).

In all retrons studied to date, it is believed that the promoter elements serve as the promoters for both *msd*RNA synthesis and the ORF. For instance, for msDNA-Ec67, promoter elements in a -10 region TTGACA and in a -35 region TGAAT, are believed to fulfill this function (see Lampson *et al.*, Science, 243, 1033-1038, 1989). However, in accordance with the invention, it is not essential that there be one promoter element for both components, but rather two promoter elements, one for initiating RNA polymerase transcription for the RT gene and the other for the *msr-msd* region. Thus the *msr-msd* region and the RT gene can be expressed under two independent promoters, which would be likely to complement each other. However, it appears at this time that at least for two of the msDNAs described herein (msDNA-Ec67 and msDNA-Ec73), the production of msDNA-Ec67 can only be complemented by the RT-Ec67 and not by the RT-Ec73 or vice-versa.

Further, it is often desired to use a strong promoter rather than the native promoter.

Another use relates to the *in vivo* production in eucaryotes of any DNA fragment(s), non-native or foreign, to the msDNA structure. Likewise, the vectors and the transfected eucaryotic hosts, carrying such foreign DNA fragment(s), the synthesis *in vivo* in eucaryotes of stable msDNAs which encompass a foreign DNA fragment in the DNA portion



or a foreign RNA fragment in the DNA portion of the DNA-RNA hybrid structure are envisaged. Of particular interest are msDNAs which include a single-strand or DNA or RNA fragment which is complementary to the mRNA of a particular target gene (or fragment) thereof (antisense DNA or RNA).

In one example of this embodiment, there was constructed a plasmid, YEp521-M5, into which there was inserted in the msd region, nucleotides 299-426 of YEp521-M4 (the boxed region of the lower strand of FIG. 7 of Lampson *et al.*, *Science*, 243, 1033-1038, 1989), a *Xho*I restriction recognition site (T↓CTAG); a foreign DNA fragment of 50-bp was inserted in this *Xho*I site. YEp521-M5 was transformed into yeast (SP-1) and the subsequently expressed msDNA designated herein as msDNA-Ye117 (see FIG. 2B).

In alike manner, there may be inserted into the msr region of YEp521-M4 a restriction recognition site, or a DNA fragment. This retron may be transformed into yeast (SP-1), and the subsequently expressed msDNA is a new structure. It corresponds to the structure designated here as msDNA-Ye117, except that the new foreign fragment is in the RNA portion of the msDNA.

Thus, the construction of a system that may be used to regulate the production of genes can be envisaged. The modified msDNAs contain in the DNA portion, a clone DNA fragment from a gene downstream of a promoter in the orientation promoting the production of antisense DNA or RNA (micRNAs). The "micRNA" terminology has been applied to an RNA transcript which is an mRNA-interfering-complementary RNA (Coleman *et al.*, *Cell*, 37, 429-436 (1984) and literature references cited therein). "micRNA" has been reported to inhibit the production of certain proteins (e.g., OmpF). A similar regulation has been reported for a micRNA and the gene for the Tn10 transposase gene. The gene for the micRNA and for the transposase are reported to be transcribed in opposite directions of the same segment of DNA, such that the 5' ends of their transcripts can form a complementary hybrid. The hybrid is thought to inhibit the translation of the transposase mRNA. Coleman *et al.*, *supra.*, report the construction of an artificial "mic" system designed to regulate the expression of any specific gene in *E. coli*.

Various cell division cycle (*cdc*) genes are known; by now some 50 different *cdc* genes have been defined in terms of landmark events occurring during duplication of cellular molecules (e.g., glycolic events). Various *cdc* genes and their functions are described in Watson *et al.*, *Molecular Biology of the Gene*, Fourth Ed. (1987), Chapter 18. Amongst these are *cdc4* required for initiation of DNA synthesis in the mitotic cell division cycle and other functions; *cdc7* of similar function to *cdc4* but for premeiotic DNA synthesis; *cdc28* necessary for duplication of the spindle pole body is homologous to mammalian protein kinases and has protein kinase activity, and others like *cdc8*, *cdc9* and others.

The strategy to produce a msDNA containing a foreign dsDNA fragment in its DNA portion is depicted in FIG. 10. The DNA fragment is shown (dark bar). The 50 bp nucleotide fragment has the following sequence (SEQ ID No.: 1):

```

5' TCGATGTAATTTCCTAATTCACCGCTCATGTTCTGAAGGATAGTTCTATTGTATC 3'
ACATTAAACGATTAAGTGGCGAGTACAAGCTTCCTATCAAGATAAACTAGAGCT 5'

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Yeast cells (SP-1) transfected with YEp521-M5 produced a new msDNA-like structure, msDNA-Ye117, shown in FIG. 2B (analyzed by polyacrylamide-urea gel electrophoresis). This new msDNA construct contains the 50-bp DNA fragment. It is contemplated that the new msDNA is a useful vector for antisense DNA. The new construct is expected to produce a single-stranded DNA which is complementary to a specific mRNA, in this instance, that of *cdc28* and inhibit the expression of that mRNA, and of the gene.

Antisense DNA (micDNA) and micRNAs which are complementary to regions of the mRNA known to interact with ribosomes, would be of particular interest. Hence, such msDNAs that contain such DNA-micDNA generating regions are of special interest for various applications. Thus, by inserting an appropriate DNA fragment of a gene after a promoter, e.g., into an *Xho*I site, one can construct with the msDNAs disclosed herein (and others) a system to specifically regulate the expression of any gene.

This is the first time that such antisense system has been provided from a molecule produced in an eucaryote.

It is contemplated that other DNA fragments be inserted in the msd region and/or the msr region of the plasmid here disclosed and the corresponding new msDNAs synthesized which may have similar functions, e.g., to generate a micDNA or a micRNA complementary to a mRNA to inhibit its gene.

Likewise, YEp521-M1 can be modified, producing an enlarged new msDNA structure (on the 5' end of the DNA portion of msDNA).

When it is desired to insert a DNA sequence encoding a protein (polypeptide) e.g., two copies of a gene, the DNA sequence will be inserted in opposite orientation to another at a selected restriction site into the msd sequence of an msDNA of choice, such as YEp521-M4. There is expected to be produced in an eucaryotic host, a novel msDNA-RNA structure. When the *lacZ* gene is incorporated into a suitable location in the msd region of the selected construct, it is

expected that  $\beta$ -galactosidase activity will be detected.

## EXAMPLES

The following Examples are offered by way of illustration and are not intended to limit the invention in any manner. In these Examples, all percentages are by weight for solids and by volume for liquids, and all temperatures are in degrees Celsius unless otherwise noted.

For convenience and clarity, the Examples refer to and provide also a detailed description of the Figures.

### Example 1

#### Yeast Strains, Media and Growth Condition

Yeast SP1 strain (a ura3 leu2 trp1 his3 ade8 can<sup>r</sup> gal2) was used. Cells were grown in YPD medium (1% yeast extract, 2% bacto-peptone, and 2% glucose). For screening transformants of YEp52 and its derivatives, a minimal medium was used (Rose *et al.*, Methods in Yeast Genetics: A Laboratory to Course Manual, Cold Spring Harbor Lab., Cold Spring Harbor, NY, 1990), supplemented with all nutrients required but leucine. For galactose induction, 0.15 ml of the pre-cultured cells in the minimal medium containing 2% galactose instead of glucose were utilized. The cells were grown at 30° until late-log phase. Yeast transformations were carried out by the lithium acetate method (6). Transformation of yeast cells was confirmed as follows: the plasmid prepared from yeast transformants was transformed into *E. coli* DH-5 (F<sup>-</sup>endA1 recA1 hsdR17 (rk<sup>-</sup>, k<sup>+</sup>) supE44 thi-1, gyrA96, relA1) and the plasmid prepared from DH-5 cells was not yet subsequently characterized. Plasmid DNA from yeast cells was prepared according to the method described by Hoffman and Winston, Gene, 57, 268-272, 1987.

Plasmids : YEp52 (Broach *et al.*, Experimental Manipulation of Gene Expression, Academic Press Inc., New York, 1983) was used to construct plasmids for expression of msDNA in yeast. This plasmid contains the ColE1 origin of replication, a promoter of the GAL10 gene, LEU2, the 2 $\mu$ -circle origin of replication, and the 2 $\mu$ -circle REP3 locus. Retron-Ec67 was prepared from plasmid pC1-1BPv4 in which the 4-kb Ba11-PvuII fragment (DNA fragment from the Ba11 to the 2nd PvuII site from the left end of the map described in FIG. 5 of Lampson *et al.*, Science, 243, 1033-1038, 1989), was cloned into the HincII site of pUC9. *E. coli* harboring this plasmid produces msDNA-Ec67. A total RNA fraction was prepared from cells transformed with pC1-1EP5c. pC1-1EP5c contains the 5-kb PstI(a)-EcoRI fragment encompassing the entire 4-kb Ba11-PvuII sequence in pC1-1BPv4 (see FIG. 5 of Lampson *et al.*, Science, 243, 1033-1038, 1989) in pUC9.

Plasmid Construction : plasmid YEp521 was constructed by introducing the multiple cloning sites of pUC19 (Yanisch-Perron *et al.*, Gene, 33, 103-119, 1985) into YEp52 (Broach *et al.*, Experimental Manipulation of Gene Expression, Academic Press Inc., New York, 1983), which was designed to obtain high-level, inducible expression of a cloned gene under the GAL10 promoter in yeast. The DNA fragment containing the pUC19 multiple cloning site was isolated by digestion of pUC19 with EcoRI; the cleaved ends were filled in with the Klenow fragment of DNA polymerase 1, and then digested with HindIII. The resulting 54-bp fragment was cloned into YEp52 by replacing a fragment between the Bc1I (filled in with the Klenow fragment) and HindIII sites, resulting in YEp521. YEp521, thus constructed, contains the multiple cloning sites from pUC19, except for EcoRI, downstream of the GAL10 promoter. The 4-kb HindIII-BamHI fragment from pC1-1BPv4 was cloned into the HindIII and BamHI sites of YEp521. As a result, the msr-msd region and the RT gene of retron-Ec67 were placed downstream of the GAL10 promoter. This plasmid is designated YEp521-M1 as shown in FIG. 7.

In order to eliminate a fragment of 242 bases upstream of msr which contains several ATG codons, polymerase chain reaction (PCR) was performed using YEp521-M1 as a template with two synthetic oligonucleotides, M2-a (5'GCAAGCTTCATAAACACGCATGT3'; SEQ ID No.: 2) and M2-b (5'CTGGATCCAGAAACGCATGCAGG3'; SEQ ID No.: 3) as primers.

These sequences correspond to the sequences from base 243 to 258 of retron Ec67 for M2-a and from base 384 to 369 for M2-b (see FIG. 7 of Watson *et al.* (22)), which flank the msr-msd region. The 140-bp PCR product was gel-purified and digested with HindIII and BamHI. The resulting fragment was cloned into the HindIII and BamHI sites of YEp521, yielding YEp521-M2. YEp521-M2 contains only the msr-msd region under the GAL10 promoter.

To insert the RT gene at the BamHI site of YEp521-M2, the 1.8-kb BamHI fragment encompassing the RT gene was amplified by PCR using YEp521-M1 as a template and two oligonucleotides, M3-a (5'CTGGATCCAGAAATGACAAAAACA3'; SEQ ID No.: 4) and M3-b (5'CTGGA TCCTT CATTG GCAT TTAAC AT3'; SEQ ID No.: 5) as primers which correspond to base 409 to 429 and from base 2182 to 2163 of retron-Ec67 (see FIG. 7 of Lampson *et al.*, Science, 243, 1033-1038, 1989), respectively. The 1.8-kb fragment was gel-purified, digested with BamHI, and closed into the BamHI site of YEp521-M2. The resulting plasmid was designated YEp521-M3.

YEp521-M4 was constructed to change the order of the msr-msd region and the RT gene. The msr-msd region

was amplified by PCR using M2-a and M2-b (see above) except that *Sma*I sites were added at their 5' ends. The 1.8-kb *Bam*HI fragment containing the RT gene was cloned into the *Bam*HI site of YE521. Subsequently, the 140-bp *Sma*I fragment containing the *msr-msd* region was cloned into the *Sma*I site of the above plasmid and the resulting plasmid was designated YE521-M4.

YE521-M5 was constructed from YE521-M4 to add the 50-bp antisense DNA for *cdc28* (*Xho*I fragment) into the *msd* region. The *Xho*I site was added into the *msd* region of YE521-M4 by PCR. This construct was then digested by *Xho*I; then the antisense DNA was ligated to the *msd* region of retron Ec67. This plasmid was transformed into yeast (SP-1) and the subsequently expressed msDNA designated herein as msDNA-Ye117.

Detection of msDNA : a total RNA fraction from yeast cells was prepared as described by Elder *et al.*, *Proc. Natl. Acad. Sci. USA*, **80**, 2432-2436, 1983 and a total RNA fraction from *E. coli* was prepared from *E. coli* harboring pC1-1EP5c by the method described by Chomzynski *et al.*, *Anal. Biochem.*, **162**, 156-159, 1987.

To label msDNA with reverse transcriptase, the total RNA fraction prepared from 0.9ml of a late-log culture was added to 20  $\mu$ l of a reaction mixture containing 30mM Tris-HCl (pH 8.2), 50mM KCl, 10mM MgCl<sub>2</sub>, 5mM DTT, 0.2mM each of dTTP, dGTP, dCTP, 5  $\mu$ Ci of [ $\gamma$ -<sup>32</sup>P]dATP and 5 units of avian myeloblastosis virus reverse transcriptase (AMV-RT; Molecular Genetic Resources). The reaction mixture was incubated at 30°C for 1 hour, and an aliquot of the reaction mixture was subjected to electrophoresis with a 6% polyacrylamide -8M urea gel. Another aliquot was treated with RNase A (10  $\mu$ g/ml) for 10 minutes at 37°C and subjected to electrophoresis.

msDNA-Ec67 was also detected by Southern blot analysis (Southern, *Mol. Biol.*, **98**, 503-517, 1975). Total RNA from 2.5ml of a late-log culture was applied to a 1.5% agarose gel with E buffer [40mM Tris HCl (pH 8.0), 10mM sodium acetate, 2mM EDTA]. After electrophoresis, the gel was blotted to a nylon membrane filter (PALL BLODYNE A TRANS-FER MEMBRANE; ICN) by the capillary transfer method. Hybridization was carried out in 50% (v/v) formamide, 5 x SSPE [1 x SSPE; 180mM NaCl, 10mM sodium phosphate (pH 7.4), 1mM EDTA], 0.3% sodium dodecyl sulfate, and 5 x Denhardt's solution (Denhardt, *Biochem. Biophys. Res. Commun.*, **23**, 641-646 (1966)) with the nick-translated 140-bp *msr-msd* region as a probe (Rigby *et al.*, *J. Mol. Biol.*, **113**, 237-251, 1977).

As noted above, the method of the invention provides for the expression of the desired msDNAs from eucaryotes. While the invention has been illustrated with a yeast of the genus *Saccharomyces*, other eucaryotes are readily suitable to practice the invention.

A convenient source of suitable yeasts is found in the *ATCC Catalogue of Yeasts*, 18th Ed., 1990. Because of practical and economic importance, the invention is particularly directed to the genus *Saccharomyces* which is extensively used in baking, beer, wine and other industries. Conventionally these yeasts are referred to as baker's, brewer's and wine yeasts.

Amongst these, of special interest are the *S. cerevisiae* strains, the *S. bayanus*, *S. carlsbergensis*, *S. diastolicus*, and *S. uvarum*, which lend themselves to transformation with the vectors of the invention. Further, to express the msDNAs, one may use vertebrate host cells like COS-1, CHO and HeLa cells or invertebrate cells or plant cells.

Plants that may be used include monocotyledons and dicotyledons. Illustrative examples of plants which may be transformed are the following: alfalfa, soybeans, maize and wheat (*Genetic Engineering of Plants, An Agricultural Perspective*, Edited by Kosuge *et al.*, Plenum Press (1983)).

Various cloning vectors may be used to transfect compatible eucaryotic host cells for replication of the vector. Thereafter the transformants are identified, plasmid DNA prepared therefrom, and the msDNAs extracted and purified.

Vectors for expression of cloned genes in yeasts are described in *Methods in Enzymology*, Vol. 194, "Guide to Yeast Genetics and Molecular Biology", page 373 (Guthrie and Fink, Eds., Academic Press Inc., 1991). It will be apparent from one skilled in the art to select an appropriate promoter for expressing msDNAs in yeast with or without a foreign DNA fragment, such as from regulatable promoters of the *GAL* family, e.g., *GAL4*, *GAL80*, *GAL1*, *GAL2*, *GAL7*, *GAL10*, *GAL11*, *MEL1*, *ADH1* and *PGK*; also see Broach *et al.*, *Experimental Manipulation of Gene Expression*, Academic Press, Inc., New York, 1983 or non-regulatable strong promoters.

Oligonucleotide synthesis may be carried out by a number of methods including those disclosed in U.S. Pat. No. 4,415,734, and in Matteucci *et al.*, *J. Am. Chem. Soc.*, **103** (11):3185-3191 (1981), Adams *et al.*, *J. Chem. Soc.*, **105** (3): 661-663 (1983), and Bemcage *et al.*, *Tetrahedron Letters*, **22** (20): (20):1859-1867 (1981).

For the expression of msDNAs in higher eucaryotes with or without selected DNA fragment, one skilled in the art may refer to and use known techniques. The advantages of synthesizing particular eucaryotic proteins in eucaryotes are well known. Depending on the msDNA which is intended to be produced, an appropriate eucaryote host cell will be selected (see *Molecular Cloning: A Laboratory Manual*, Second Edition, §3, §16.3 and seq. (Sambrook *et al.*, Cold Spring Harbor Laboratory Press, 1989)). The eucaryotic expression vehicle will contain, as is known, a promoter and enhancer elements, recognition sequences, the TATA box and upstream promoter elements. Other conventional elements located upstream of the transcription initiation site for replication and selection are known and described in standard laboratory manuals. Vectors are available commercially, for instance from Pharmacia (pMSG, pSVT17, pMT2). For methods for introducing recombinant vectors into mammalian cells, see *Molecular Cloning: A Laboratory Manual*, Second Edition, §16.30-16.55, (Sambrook *et al.*, Cold Spring Harbor Laboratory Press, 1989). For cosmid

vectors for transfection of mammalian cells, see Molecular Cloning: A Laboratory Manual, Second Edition, \$23.18 and seq., (Sambrook et al., Cold Spring Harbor Laboratory Press, 1989).

Further, one skilled in the art may wish to refer to Current Protocols In Molecular Biology, Volume 1, §16. 12-16. 13.7 (Ausubel et al., Eds., , Greene Publishing Associates and Wiley-Interscience, 1989), discussing in particular, three vector systems or strategies for introducing foreign genes into mammalian cells with COS cells, CHC and vaccinia viral vectors. One skilled in the art will select the most appropriate system for the production of msDNAs from the selected retrans, and further, for introduction of DNA into mammalian cells (see Current Protocols In Molecular Biology, Volume 1, §9. 01-9.93 (Ausubel et al., Eds., , Greene Publishing Associates and Wiley-Interscience, 1989)).

The msDNAs have several interesting utilities.

A fascinating utility that is being considered is the role that msDNAs can play on the formation of triple helix DNA, or triplex DNA with a specific duplex on the chromosome. A recent report in Science, 252, 1374-1375 (June 27, 1991), "Triplex DNA Finally Comes of Age", highlights the timeliness of the present invention. Triplex DNA can be formed by binding a third strand to specific recognized sites on chromosomal DNA. Synthetic strands of sizes preferably containing the full complement of bases (such as 11-15 and higher), are discussed. The msDNAs with long 3'(or 5') ends (and the loop of non-duplexed bases) would appear to be excellent candidates. These regions provide single-stranded DNA necessary for the triplex formation. The resulting triplex DNA is expected to have increased stability and usefulness. New therapies based on the triple helix formation, including in AIDS therapy and selective gene inhibition and others are proposed in the Report.

Artificial, synthetic msDNAs can be designed and may be used as antisense DNAs, and/or RNAs and/or ribozymes using the single-stranded DNA or RNA region of msDNAs. Such msDNA (containing a foreign ssDNA or ssRNA fragment) for use as antisense system, been described above. The production of an msDNA with complementarity with a gene (or portion) thereof, blocks the synthesis of the specific protein itself. The msDNA system produced in eucaryotic cells to generate a desired complementary DNA of an mRNA of a gene, appears to have real potential in eucaryotic cells to block the expression of various harmful or toxic genes, such as drug resistance, oncogenes, and phages or viruses. The system could have applications to AIDS therapy. Of special interest, are the msDNAs that would be produced by HeLa cells and containing such selected DNA fragment for use in antisense applications.

As described above, it is contemplated that genes be inserted for instance, in the the stem region(s) of the msDNAs. Thus the msDNAs may be used for amplification of the selected gene.

The polymerase chain reaction (PCR) is a well-known rapid procedure for in vitro enzymatic amplification of a specific segment of DNA. The standard PCR method requires a segment of double-stranded DNA to be amplified, and always two-single stranded oligonucleotide primers flanking the segment, a DNA polymerase, appropriate deoxyribonucleoside triphosphate (dNTPs), a buffer, and salts (Current Protocols, Section 15).

Thus, the msDNAs due to their unique structure (and stability), are expected to be of value in numerous applications in the biochemical, medical, pharmaceutical and other biological sciences.

TABLE I  
Summary of the structure of *mdm3*

	Structure of <i>mdm3</i> <sup>a</sup>					Reversed transcription		
	Length of <i>mdm3</i> (nt)	Length of <i>mdm3</i> (nt)	3' end overlap length (nt)	Inverted repeat length (nt)	Position of the branched G	Copy number per cell <sup>c</sup>	WT ORF and RT ORF	Distances between and RT ORF
Hx162	182	77	8	34	5'-20	500-700	485	77
Hx65	65	49 (62) <sup>d</sup>	6	15	6'-4	100	427	28
St163	163	76	8	13	(6'-17) <sup>e</sup>	500	ND	ND
Bc67	67	58	7	13	6'-15	500	586	51
Bc86	86	82	11	12	6'-14	500	120	19
Bc73	73	75 <sup>f</sup>	5	13	6'-13	ND	316	53
Bc107	107	75	6	16	6'-18	ND	319	50
				</				

<sup>a</sup> See Figure 2.

<sup>b</sup> The length of the a1 and a2.

<sup>c</sup> Copy numbers are estimated approximately.

<sup>d</sup> On the basis of the inverted repeat structures, the primary product is considered to have a longer 5' arm of 13 bases.

<sup>e</sup> The distance between and the first cut. The 61 base overlaps by 4 codons (Sun et al., *Science*, submitted (1991)).

<sup>f</sup> On the basis of the inverted repeat structures, the lengths of the 5' arm were estimated to be 16, 14, and 17 bases for Hx65, Bc73 and Bc107, respectively.

<sup>g</sup> Okada et al., *Cell*, 51, 1105-12 (1987) and Ren et al., *Cell*, 29, 203-9 (1984).

<sup>h</sup> Okada et al., *Cell*, 51, 1101. *Open*, 25, 9055-58 (1988).

<sup>i</sup> Purich et al., *Cell*, 49, 47-53 (1987) and Purich et al., *Cell*, 49, 55-62 (1987).

<sup>j</sup> Longman et al., *Science*, 243, 1033-36 (1989).

<sup>k</sup> Liu and Hsiao, *Cell*, 56, 891-904 (1989).

<sup>l</sup> Sun et al., *Science*, submitted (1991).

<sup>m</sup> Herzer et al., *Mol. Microbiol.*, submitted (August 1991).

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## SEQUENCE LISTING

## (1) GENERAL INFORMATION:

(i) APPLICANT: University of Medicine and Dentistry of New Jersey

(ii) TITLE OF INVENTION: Method for synthesizing stable single-stranded cDNA in eukaryotes by means of a bacterial retron, products and uses therefor.

(iii) NUMBER OF SEQUENCES: 5

(iv) CORRESPONDENCE ADDRESS:

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(B) STREET: 55, Rue d'Amsterdam  
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(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk  
(B) COMPUTER: IBM PC compatible  
(C) OPERATING SYSTEM: PC-DOS/MS-DOS  
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(vii) PRIOR APPLICATION DATA:

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(viii) ATTORNEY/AGENT INFORMATION:

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(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: 42 80 64 68  
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(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 54 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: double  
(D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: DNA (genomic)

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

TCGATGTAAT TTGCTAATTC ACCGCTCATG TTCGAAGGAT AGTTCTATTT GATC

54

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 23 base pairs

- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

5 (ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

10

GCAAGCTTCA TAAACACGCA TGT

23

15 (2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 23 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

20

(ii) MOLECULE TYPE: cDNA

25

(iii) HYPOTHETICAL: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

30

CTGGATCCAG AAACGCATGC AGG

23

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

35

(ii) MOLECULE TYPE: cDNA

40

(iii) HYPOTHETICAL: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

45

CTGGATCCAA GAAATGACAA AAACA

25

50

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

55



(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

CTGGATCCCT CATTAGCTAT TTAACAT

27

## Claims

1. A method for producing a stable msDNA in a eucaryotic host cell which comprises transfecting the cell with a DNA expression vector which replicates in the cell, which vector contains a retron encoding the msDNA, which retron contains msr and msd coding regions (msr-msd) for the msDNA and a gene encoding a reverse transcriptase (RT), and expressing the msDNA from the retron.
2. The method of claim 1 wherein the expression vector is a plasmid.
3. The method of claim 2 wherein the eucaryote host cell is yeast.
4. The method of claim 3 wherein the yeast is of the genus Saccharomyces.
5. The method of claim 1 wherein the eucaryote host cell is a plant or mammalian cell.
6. The method of claim 3 wherein in the retron, the RT gene is downstream of the msr-msd region.
7. The method of claim 3 wherein the retron contains a single AUG codon for the RT gene.
8. The method of claim 7 wherein in the retron, the RT gene is upstream of the msr-msd region.
9. The method of claim 3 wherein the expression vector contains a promoter for a retron consisting of msr-msd region and a RT gene, which promoter is located upstream of msr.
10. The method of claim 9 wherein the promoter is a foreign strong promoter.
11. The method of claim 10 wherein the strong promoter is GAL10.
12. The method of claim 2 wherein in the retron in the plasmid, the msd sequence contains a cloning site.
13. The method of claim 8 wherein a foreign DNA fragment is contained in the msd region of the retron which codes the DNA of the msDNA.
14. The method according to anyone of claims 1-13 wherein the retron is in a vector selected from the group consisting of YEp521-M1 (ATCC N° 74092), YEp521-M3, YEp521-M4 (ATCC N° 74093), and YEp521-M5 (ATCC N° 74094).
15. The method of claim 4 wherein the msDNA molecule comprises a branched single-stranded RNA which is covalently linked to a single-stranded DNA by a 2',5'-phosphodiester bond between the 2'-OH group and an internal rG residue and the 5'-phosphate of the DNA molecule, which RNA is non-covalently linked to the DNA by base-pairing between the complementary 3' ends of the RNA and DNA molecules, which RNA-DNA form stable stem-loop secondary structures, which msDNA is encoded by a primary RNA transcript, pre-msDNA which contains an open-reading frame (ORF) downstream of the msr locus, the ORF encoding a polypeptide which has sequence similarity with retroviral RTs and a highly conserved sequence common to all RTs.
16. The method of claim 15 wherein the stable msDNA which is produced is selected from the group consisting of msDNA-Ec67, msDNA-h4x162, msDNA-Mx65, msDNA-Ec107, msDNA-Mx86 and msDNA-Sal63, as shown in Figure 2A.

17. The method of claim 4 wherein the ORF is located upstream of the msr locus.
18. The method of claim 17 wherein the stable msDNA which is produced is msDNA-Ec67, as shown in Figure 2A.
- 5 19. A DNA expression vector which replicates in an eucaryotic host which comprises a retron encoding a stable hybrid single-stranded RNA-DNA (msDNA) structure, which retron contains a gene encoding a reverse transcriptase (RT) and one coding region which contains two coding sequences, one "msr" and the other "msd" for encoding, respectively, the RNA and DNA portions of the msDNA.
- 10 20. The DNA expression vector of claim 19 wherein in the retron, the RT gene is downstream of the msr-msd region.
21. The vector of claim 20 wherein the retron contains a single AUG codon for the RT gene.
22. The vector of claim 21 wherein in the retron, the RT gene is upstream of the msr-msd region.
- 15 23. The vector of claim 19 which includes a promoter for the msr-msd region and the RT gene, which promoter is located upstream of msr.
24. The vector of claim 23 wherein the promoter is a foreign strong promoter.
- 20 25. The vector of claim 24 wherein the promoter is GAL10.
26. The vector of claim 22 in which a foreign DNA fragment is contained in the msd region of the retron which encodes the DNA of the msDNA.
- 25 27. The vector of claim 26 wherein the fragment is 50-bp long.
28. The vector of claim 20 which is YEp521-M1 (ATCC N° 74092), YEp521-M3, YEp521-M4 (ATCC N° 74093) or YEp521-M5 (ATCC N° 74094).
- 30 29. A eucaryotic host cell transfected with a DNA expression vector which replicates in the cell, which vector contains a retron encoding a msDNA, which retron contains msr and msd coding regions (msr-msd) for the msDNA and a gene encoding a reverse transcriptase (RT).
- 35 30. The eucaryotic host cell of claim 29 which is selected from the group consisting of yeast, plant cells and mammalian cells.
31. The eucaryotic host cell of claim 30 which is yeast.
- 40 32. The eucaryotic host cell of claim 31 wherein the yeast is of the genus Saccharomyces.
33. The eucaryotic host cell of claim 30, 31 or 32 wherein in the retron, the RT gene is downstream of the msr-msd region.
- 45 34. The eucaryotic host cell of claim 30 wherein the retron contains a single AUG codon for the RT gene.
35. The eucaryotic host cell of claim 34 wherein in the retron, the RT gene is upstream of the msr-msd region.
- 50 36. The eucaryotic host cell of claim 30 wherein the expression vector contains the promoter for the msr-msd region and RT gene, which promoter is located upstream of msr.
37. The eucaryotic host cell of claim 36 wherein the promoter is a foreign strong promoter.
- 55 38. The eucaryotic host cell of claim 37 wherein the strong promoter is GAL10.
39. The eucaryotic host cell of claim 35 which contains a retron having a msd region which contains a foreign DNA fragment.

40. The eucaryotic host cell of claim 39 wherein the DNA sequence is 50-bp long.

# Patentansprüche

1. Verfahren zur Herstellung einer beständigen ms-DNA in einer eukaryotischen Wirtszelle, das die Transfektion der Zelle mit einem DNA-Expressionsvektor, der in der Zelle repliziert, und ein die ms-DNA kodierendes Retron enthält, das eine msr- und eine msd-Kodierungsregion (msr-msd) für die ms-DNA sowie ein eine Reverse-Transcriptase (RT) kodierendes Gen enthält, und die Expression der ms-DNA aus dem Retron umfaßt.
2. Verfahren nach Anspruch 1, bei dem der Expressionsvektor ein Plasmid ist.
3. Verfahren nach Anspruch 2, bei dem die eukaryotische Wirtszelle Hefe ist.
4. Verfahren nach Anspruch 3, bei dem die Hefe der Gattung *Saccharomyces* angehört.
5. Verfahren nach Anspruch 1, bei dem die eukaryotische Wirtszelle eine Pflanzen- oder Säugerzelle ist.
6. Verfahren nach Anspruch 3, bei dem im Retron das RT-Gen unterhalb der msr-msd-Region angeordnet ist.
7. Verfahren nach Anspruch 3, bei dem das Retron ein einziges AUG-Kodon für das RT-Gen enthält.
8. Verfahren nach Anspruch 7, bei dem im Retron das RT-Gen oberhalb der msr-msd-Region angeordnet ist.
9. Verfahren nach Anspruch 3, bei dem der Expressionsvektor einen Promoter für ein aus der msr-msd-Region und einem RT-Gen bestehendes Retron enthält und der Promoter oberhalb der msr-Region angeordnet ist.
10. Verfahren nach Anspruch 9, bei dem der Promoter ein fremder starker Promoter ist.
11. Verfahren nach Anspruch 10, bei dem der starke Promoter GAL 10 ist.
12. Verfahren nach Anspruch 2, bei dem im Retron im Plasmid die msd-Sequenz einen Klonungssite enthält.
13. Verfahren nach Anspruch 8, bei dem ein fremdes DNA-Fragment in der msd-Region des Retron enthalten ist, das die DNA der ms-DNA kodiert.
14. Verfahren nach einem der Ansprüche 1 bis 13, bei dem das Retron in einem Vektor enthalten ist, ausgewählt aus der Gruppe, bestehend aus YEp521-M1 (ATCC Nr. 74092), YEp521-M3, YEp521-M4 (ATCC Nr. 74093) und YEp521-M5 (ATCC Nr. 74094).
15. Verfahren nach Anspruch 4, bei dem das ms-DNA-Molekül eine verzweigte einsträngige RNA umfaßt, die mit einer einsträngigen DNA durch eine 2',5'-Phosphodiester-Bindung zwischen der 2'-OH-Gruppe von einem inneren rG-Rest und dem 5'-Phosphat des DNA-Moleküls kovalent verknüpft ist und mit der DNA durch Basenpaarung zwischen den komplementären 3'-Enden der RNA- und DNA-Moleküle nicht-kovalent verknüpft ist, die RNA-DNA beständige sekundäre Stem-Loop-Strukturen bilden, die ms-DNA durch ein primäres RNA-Transcript, die prä ms-DNA, kodiert wird, die einen offenen Leserahmen (ORF) unterhalb des msr-Locus enthält, der ein Polypeptid kodiert, das mit retroviralen RT's Sequenzähnlichkeit aufweist und eine mit sämtlichen RT's gemeinsame, in hohem Maße konservierte Sequenz umfaßt.
16. Verfahren nach Anspruch 15, bei dem die beständige ms-DNA, die produziert wird, aus der Gruppe, bestehend aus ms-DNA-Ec67, ms-DNA-Mx162, ms-DNA-Mx65, ms-DNA-Ec107, ms-DNA-Mx86 und ms-DNA-Sa163, wie in Fig. 2A gezeigt, ausgewählt wird.
17. Verfahren nach Anspruch 4, bei dem der ORF oberhalb des msr-Locus angeordnet ist.
18. Verfahren nach Anspruch 17, bei dem die beständige ms-DNA, die produziert wird, wie in Fig. 2A gezeigt, ms-DNA-Ec67 ist.

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19. DNA-Expressionsvektor, der in einem eukaryotischen Wirt repliziert, der ein Retron umfaßt, das eine beständige einsträngige Hybrid-RNA-DNA (ms-DNA)-Struktur kodiert und ein Gen, das eine Reverse-Transcriptase (RT) kodiert, sowie eine Kodierungsregion enthält, die zwei Kodierungssequenzen umfaßt, und zwar eine msr-Sequenz für die Kodierung des RNA-Anteils und eine msd-Sequenz für die Kodierung des DNA-Anteils der ms-DNA.
20. DNA-Expressionsvektor nach Anspruch 19, bei dem im Retron das RT-Gen unterhalb der msr-msd-Region angeordnet ist.
21. Vektor nach Anspruch 20, bei dem das Retron ein einziges AUG-Kodon für das RT-Gen enthält.
22. Vektor nach Anspruch 21, bei dem im Retron das RT-Gen oberhalb der msr-msd-Region angeordnet ist.
23. Vektor nach Anspruch 19, der einen Promoter für die msr-msd-Region und das RT-Gen umfaßt und der Promoter oberhalb der msr-Region angeordnet ist.
24. Vektor nach Anspruch 23, bei dem der Promoter ein fremder starker Promoter ist.
25. Vektor nach Anspruch 24, bei dem der Promoter GAL10 ist.
26. Vektor nach Anspruch 22, bei dem ein fremdes DNA-Fragment in der msd-Region des Retron enthalten ist, das die DNA der ms-DNA kodiert.
27. Vektor nach Anspruch 26, bei dem das Fragment 50 bp lang ist.
28. Vektor nach Anspruch 20, der YEp521-M1 (ATCC Nr. 74092), YEp521-M3, YEp521-M4 (ATCC Nr. 74093) oder YEp521-M5 (ATCC Nr. 74094) darstellt.
29. Eukaryotische Wirtszelle, die mit einem DNA-Expressionsvektor transfiziert ist, der in der Zelle repliziert, wobei der Vektor ein eine ms-DNA kodierendes Retron enthält, das eine msr- und eine msd-Kodierungsregion (msr-msd) für die ms-DNA sowie ein eine Reverse-Transcriptase (RT) kodierendes Gen enthält.
30. Eukaryotische Wirtszelle nach Anspruch 29, die aus der Gruppe, bestehend aus Hefe, Pflanzen- und Säugerzellen, ausgewählt wird.
31. Eukaryotische Wirtszelle nach Anspruch 30, die Hefe darstellt.
32. Eukaryotische Wirtszelle nach Anspruch 31, bei der die Hefe der Gattung *Saccharomyces* angehört.
33. Eukaryotische Wirtszelle nach Anspruch 30, 31 oder 32, bei der im Retron das RT-Gen unterhalb der msr-msd-Region angeordnet ist.
34. Eukaryotische Wirtszelle nach Anspruch 30, bei der das Retron ein einziges AUG-Kodon für das RT-Gen enthält.
35. Eukaryotische Wirtszelle nach Anspruch 34, bei der im Retron das RT-Gen oberhalb der msr-msd-Region angeordnet ist.
36. Eukaryotische Wirtszelle nach Anspruch 30, bei der der Expressionsvektor den Promoter für die msr-msd-Region und das RT-Gen enthält und der Promoter oberhalb der msr-Region angeordnet ist.
37. Eukaryotische Wirtszelle nach Anspruch 36, bei der der Promoter ein fremder starker Promoter ist.
38. Eukaryotische Wirtszelle nach Anspruch 37, bei der der starke Promoter GAL10 ist.
39. Eukaryotische Wirtszelle nach Anspruch 35, die ein Retron mit einer msd-Region enthält, die ein fremdes DNA-Fragment enthält.
40. Eukaryotische Wirtszelle nach Anspruch 39, bei der die DNA-Sequenz 50 bp lang ist.

# Revendications

1. Procédé de production d'un ADNms stable dans une cellule hôte eucaryote, qui comprend la transfection de la cellule avec un vecteur d'expression d'ADN qui se réplique dans la cellule, ce vecteur contenant un rétron codant pour l'ADNms, lequel rétron contient les régions codantes *msr* et *msd* (*msr-msd*) pour l'ADNms et un gène codant pour une transcriptase inverse (RT), et l'expression de l'ADNms à partir du rétron.
2. Procédé selon la revendication 1, dans lequel le vecteur d'expression est un plasmide.
3. Procédé selon la revendication 2, dans lequel la cellule hôte eucaryote est une levure.
4. Procédé selon la revendication 3, dans lequel la levure est du genre *Saccharomyces*.
5. Procédé selon la revendication 1, dans lequel la cellule hôte eucaryote est une cellule végétale ou une cellule de mammifère.
6. Procédé selon la revendication 3, dans lequel, dans le rétron, le gène de RT est en aval de la région *msr-msd*.
7. Procédé selon la revendication 3, dans lequel le rétron contient un seul codon AUG pour le gène de RT.
8. Procédé selon la revendication 7, dans lequel, dans le rétron, le gène de RT est en amont de la région *msr-msd*.
9. Procédé selon la revendication 3, dans lequel le vecteur d'expression contient un promoteur pour un rétron constitué d'une région *msr-msd* et d'un gène de RT, ce promoteur étant situé en amont de *msr*.
10. Procédé selon la revendication 9, dans lequel le promoteur est un promoteur fort étranger.
11. Procédé selon la revendication 10, dans lequel le promoteur fort est *GAL10*.
12. Procédé selon la revendication 2, dans lequel, dans le rétron contenu dans le plasmide, la séquence *msd* contient un site de clonage.
13. Procédé selon la revendication 8, dans lequel un fragment d'ADN étranger est contenu dans la région *msd* du rétron qui code pour l'ADN de l'ADNms.
14. Procédé selon l'une quelconque des revendications 1 à 13, dans lequel le rétron est contenu dans un vecteur choisi dans le groupe constitué par YEp521-M1 (ATCC n° 74092), YEp521-M3, YEp521-M4 (ATCC n° 74093) et YEp521-M5 (ATCC n° 74094).
15. Procédé selon la revendication 4, dans lequel la molécule d'ADNms comprend un ARN simple brin ramifié, lié par covalence à un ADN simple brin par une liaison phosphodiester 2',5' entre le groupe 2'-OH d'un résidu rG interne et le phosphate en 5' de la molécule d'ADN, cet ARN est lié de manière non covalente à l'ADN par un appariement de bases entre les extrémités complémentaires 3' des molécules d'ARN et d'ADN, ce système ARN-ADN forme des structures secondaires de type tige-boucle stables, cet ADNms est codé par un transcrit ARN primaire, un pré-ADNms qui contient un cadre de lecture ouvert (ORF) en aval du locus *msr*, l'ORF codant pour un polypeptide qui a une analogie de séquence avec des RT rétroviraux et une séquence hautement conservée commune à toutes les RT.
16. Procédé selon la revendication 15, dans lequel l'ADNms stable qui est produit est choisi dans le groupe constitué par l'ADNms-Ec67, l'ADNms-Mx162, l'ADNms-Mx65, l'ADNms-Ec107, l'ADNms-Mx86 et l'ADNms-Sal63, tels que présentés dans la figure 2A.
17. Procédé selon la revendication 4, dans lequel l'ORF est situé en amont du locus *msr*.
18. Procédé selon la revendication 17, dans lequel l'ADNms stable qui est produit est l'ADNms-Ec67, tel que présenté dans la figure 2A.
19. Vecteur d'expression d'ADN qui se réplique dans un hôte eucaryote et qui comprend un rétron codant pour une

structure ARN-ADN simple brin hybride stable (ADNms), ce rétron contenant un gène codant pour une transcriptase inverse (RT) et une région codante contenant deux séquences codantes, l'une *msr* et l'autre *msd*, codant respectivement pour les portions d'ARN et d'ADN de l'ADNms.

20. Vecteur d'expression d'ADN selon la revendication 19, dans lequel dans le rétron, le gène de RT est en aval de la région *msr-msd*.
21. Vecteur selon la revendication 20, dans lequel le rétron contient un seul codon AUG pour le gène de RT.
22. Vecteur selon la revendication 21, dans lequel, dans le rétron, le gène de RT est en amont de la région *msr-msd*.
23. Vecteur selon la revendication 19, qui comprend un promoteur pour la région *msr-msd* et le gène de RT, ce promoteur étant situé en amont de *msr*.
24. Vecteur selon la revendication 23, dans lequel le promoteur est un promoteur fort étranger.
25. Vecteur selon la revendication 24, dans lequel le promoteur est *GAL10*.
26. Vecteur selon la revendication 22, dans lequel un fragment d'ADN étranger est contenu dans la région *msd* du rétron qui code pour l'ADN de l'ADNms.
27. Vecteur selon la revendication 26, dans lequel le fragment a une longueur de 50 bp.
28. Vecteur selon la revendication 20, qui est YEp521-M1 (ATCC n° 74092), YEp521-M3, YEp521-M4 (ATCC n° 74093) ou YEp521-M5 (ATCC n° 74094).
29. Cellule hôte eucaryote transfectée avec un vecteur d'expression d'ADN qui se réplique dans la cellule, ce vecteur contenant un rétron codant pour un ADNms, ce rétron contenant des régions codantes *msr* et *msd* (*msr-msd*) pour l'ADNms et un gène codant pour une transcriptase inverse (RT).
30. Cellule hôte eucaryote selon la revendication 29, qui est choisie dans le groupe constitué par des levures, des cellules végétales et des cellules de mammifère.
31. Cellule hôte eucaryote selon la revendication 30, qui est une levure.
32. Cellule hôte eucaryote selon la revendication 31, dans laquelle la levure est du genre *Saccharomyces*.
33. Cellule hôte eucaryote selon la revendication 30, 31 ou 32, dans laquelle, dans le rétron, le gène de RT est en aval de la région *msr-msd*.
34. Cellule hôte eucaryote selon la revendication 30, dans laquelle le rétron contient un seul codon AUG pour le gène de RT.
35. Cellule hôte eucaryote selon la revendication 34, dans laquelle, dans le rétron, le gène de RT est en amont de la région *msr-msd*.
36. Cellule hôte eucaryote selon la revendication 30, dans laquelle le vecteur d'expression contient le promoteur pour la région *msr-msd* et le gène de RT, ce promoteur étant situé en amont de *msr*.
37. Cellule hôte eucaryote selon la revendication 36, dans laquelle le promoteur est un promoteur fort étranger.
38. Cellule hôte eucaryote selon la revendication 37, dans laquelle le promoteur fort est *GAL10*.
39. Cellule hôte eucaryote selon la revendication 35, qui contient un rétron ayant une région *msd* contenant un fragment d'ADN étranger.
40. Cellule hôte eucaryote selon la revendication 39, dans laquelle la séquence d'ADN a une longueur de 50 bp.

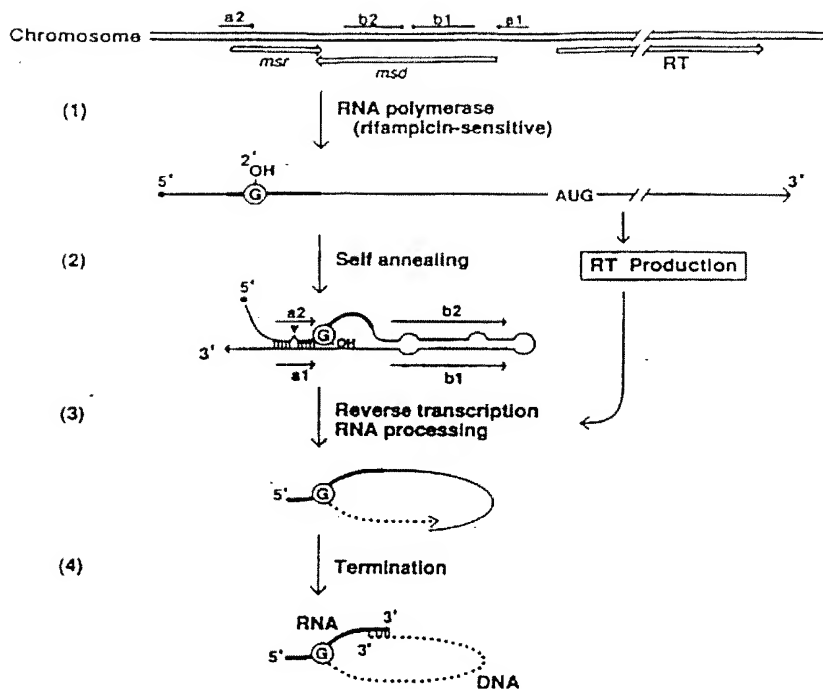


FIG.1

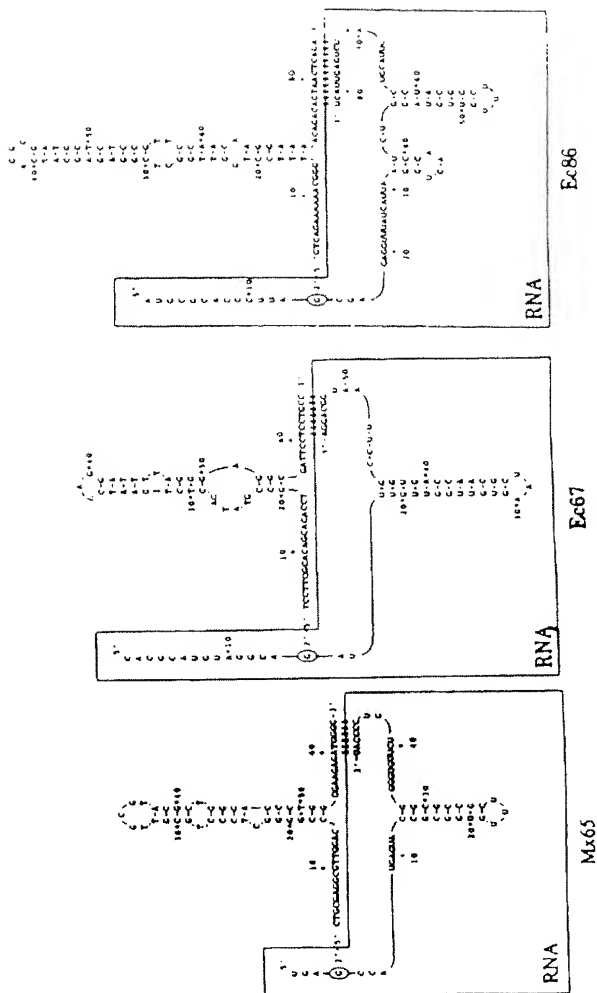


FIG.2A



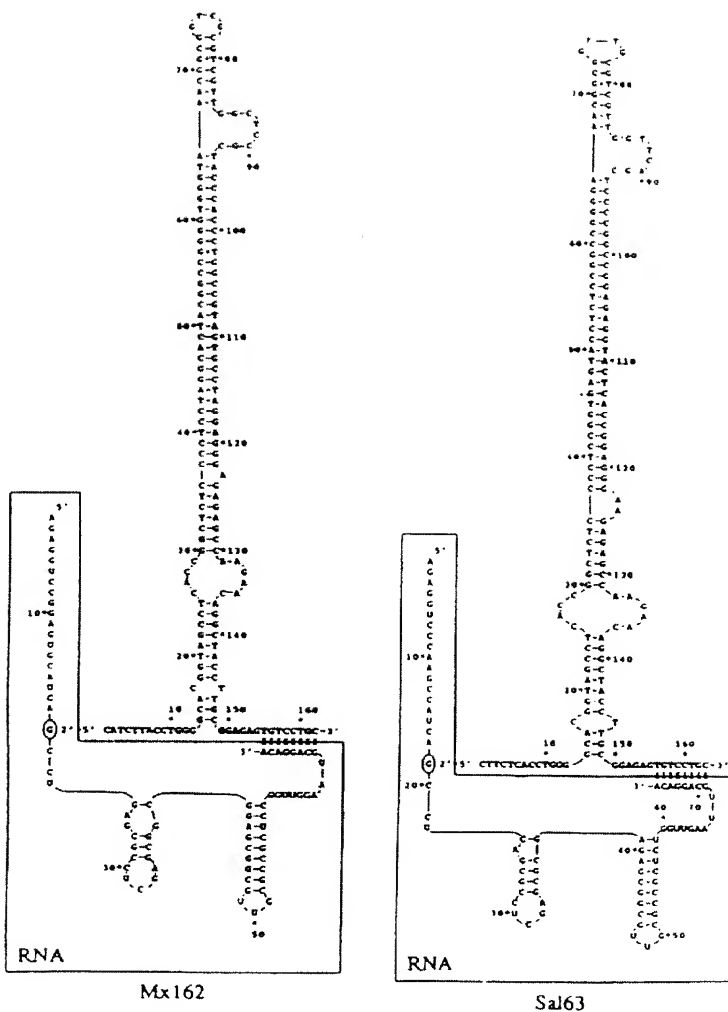


FIG.2A(continued)

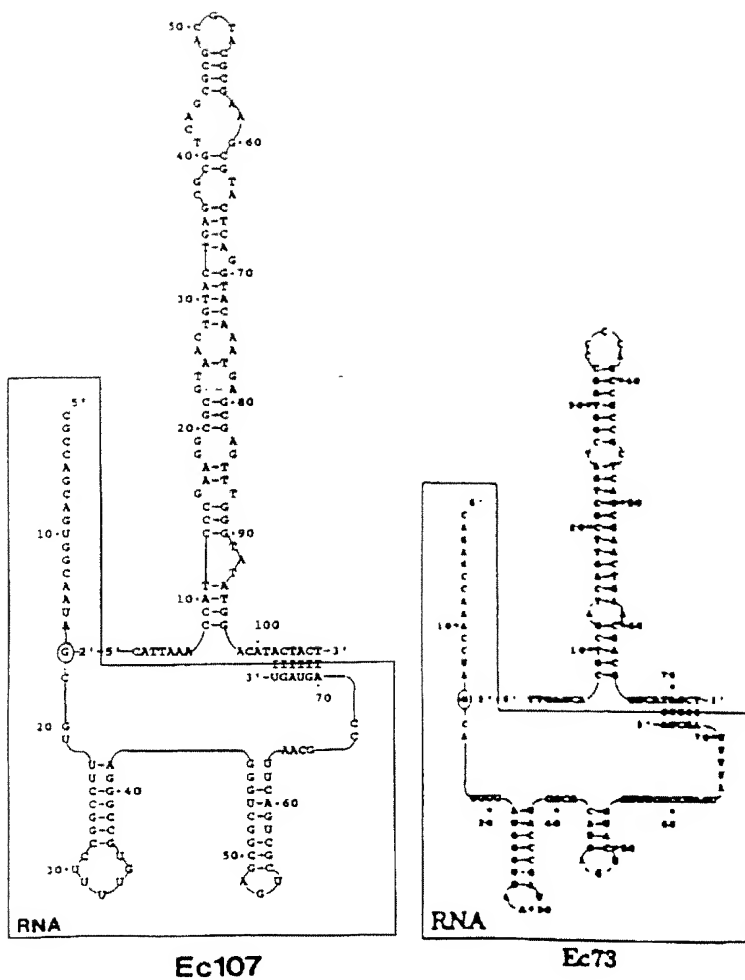


FIG.2A(continued)

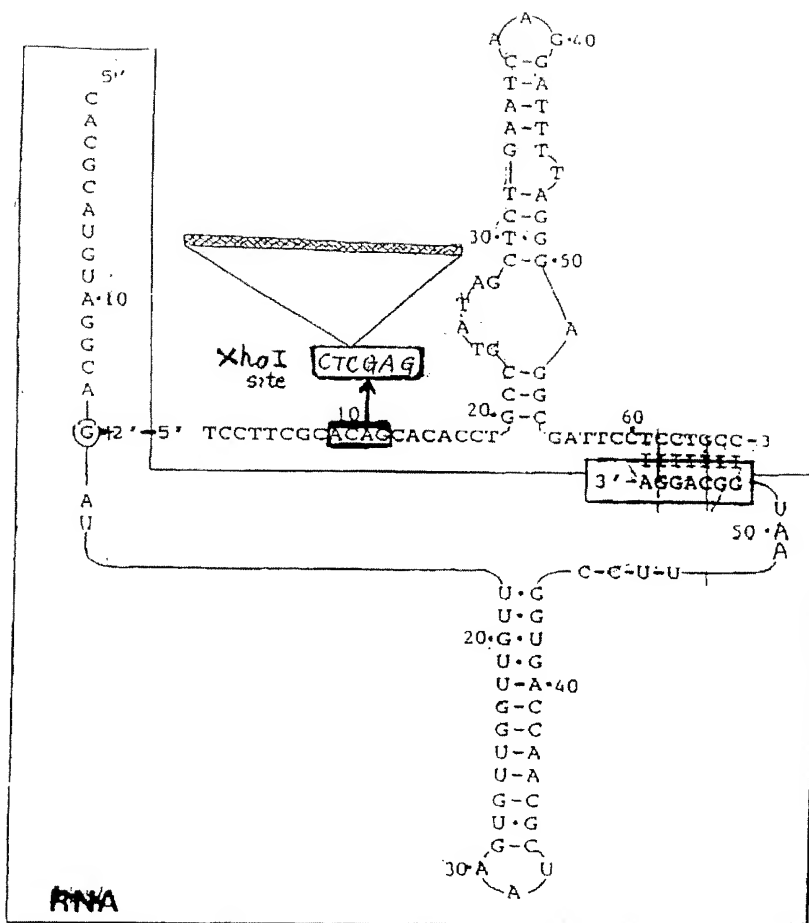


FIG. 2B

8

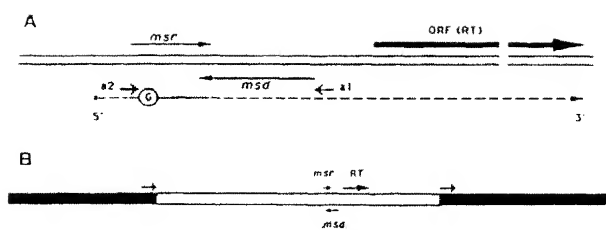


FIG.3

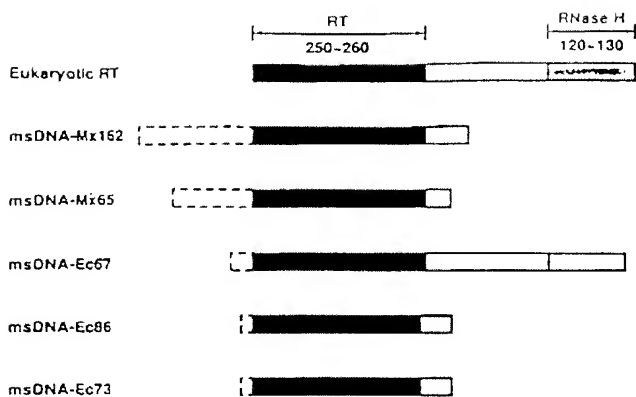


FIG.4

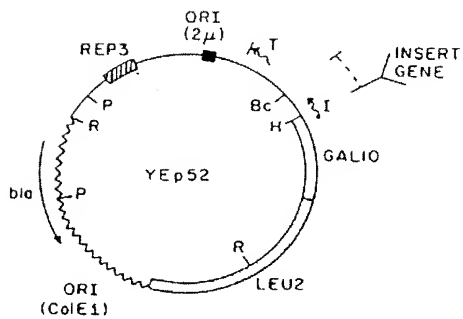
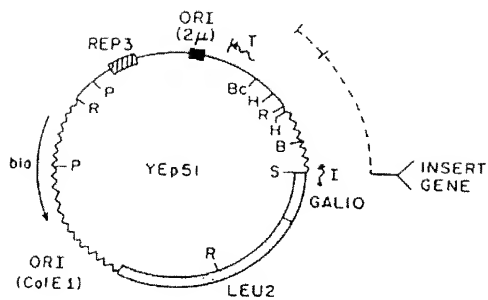


FIG.5

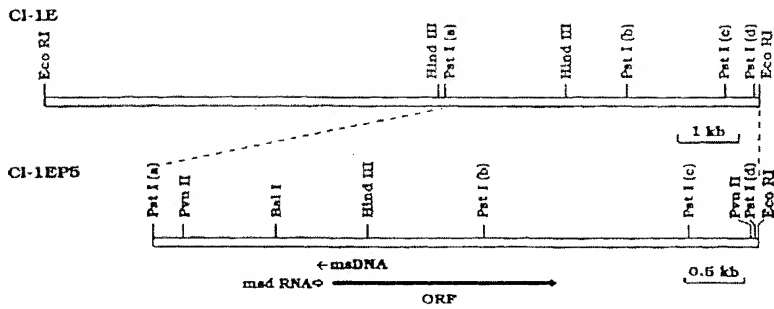


FIG.6

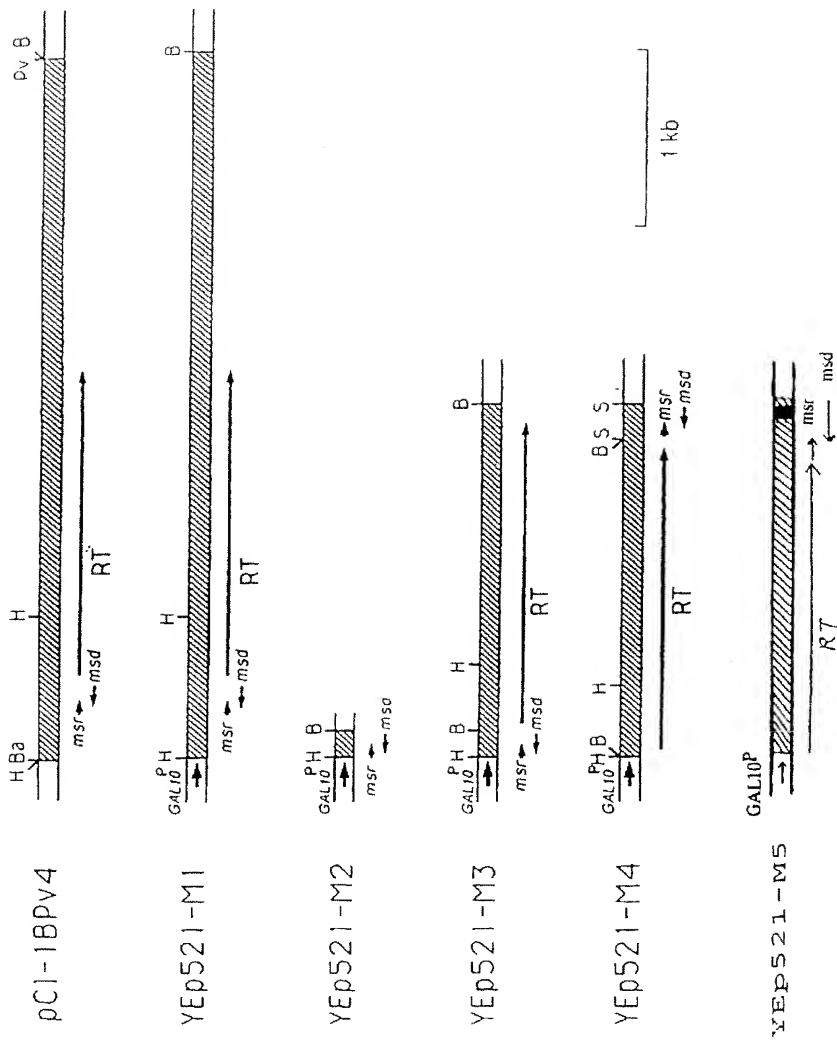


FIG.7



123456

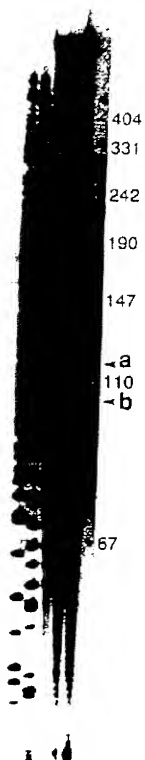


FIG.8A

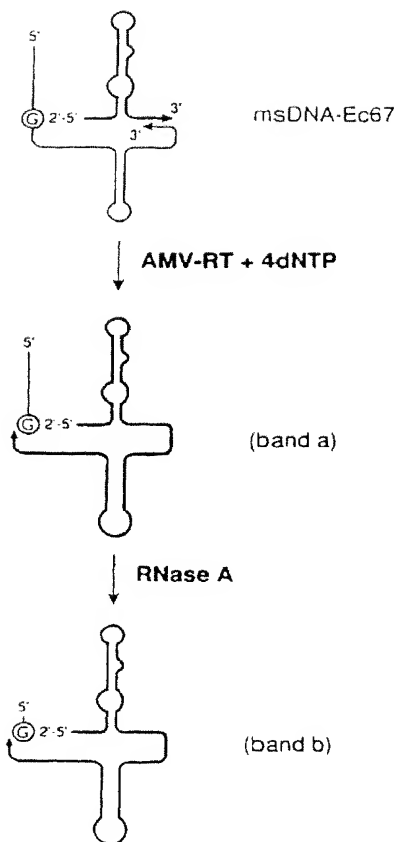


FIG.8B

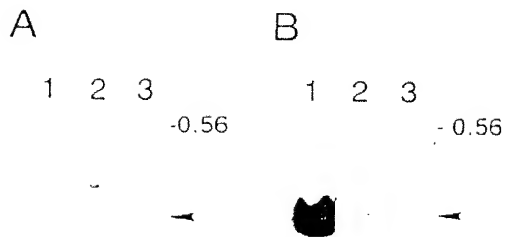


FIG.9

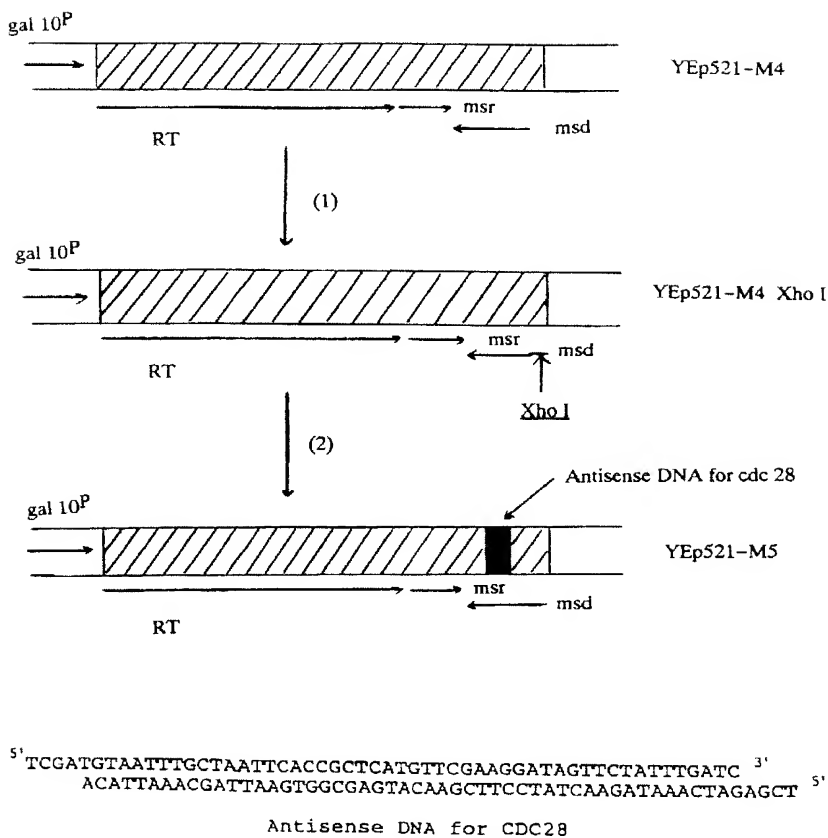


FIG.10